

Introduction to Gene Set Testing and the Global Test

Utah State University – Spring 2014
STAT 5570: Statistical Bioinformatics
Notes 4.3

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References

- Goeman et al. Bioinformatics 20(1):93-99 (2004)
- Goeman et al. JRSS-B 68(3) 477-493 (2006)
- Goeman & Buhlmann. Bioinformatics 23(8):980-987 (2007)
- Chapters 19-22 of Bioconductor Monograph (course text)

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Motivation for “over-representation”

- Suppose we perform a test of DE and find a list of 132 significant genes (out of 2,000)
- Consider a specific GO term, like apoptosis

	apop.	not apop.	
significant	100	32	132
not signif.	753	1,115	1,868
	853	1,147	2,000

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Questions in Gene Set Tests

- Is “significance” independent of “apoptosis”?
- Is the gene set “apoptosis” over-represented among “significant” genes?
- Is the gene set “apoptosis” differentially expressed?

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Traditional test of “independence”

- EV = (row total) x (column total) / (table total)
- Test statistic:

$$\chi^2 = \sum \frac{(\text{OV} - \text{EV})^2}{\text{EV}}$$

- If truth is “independence” and sample size is large, then $\chi^2 \sim \chi_{df}^2$ $df = (\# \text{ cols.} - 1)(\# \text{ rows} - 1)$
- Or – obtain P-value by permutations (exact test, based on hypergeometric dist'n)

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Hypergeometric distn. & Fisher's exact test

- Suppose an urn contains n balls:
 r black and $n-r$ white
- Draw m balls without replacement
- Let $X = \#$ black balls drawn;
then $X \sim \text{hypergeometric}(r, n, m)$:

$$P(X = k) = \frac{\binom{r}{k} \binom{n-r}{m-k}}{\binom{n}{m}}$$

- Use this to generate probabilities of all possible 2x2 tables with same row and column totals; look at probability of first cell count
- P-value = proportion of tables more extreme than the original table (usually based on χ^2 values)

(implemented in hyperGTest function of package GOstats)

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But wait – possible null hypotheses

- Competitive null

compare DE of gene set (G) to a standard defined by the set's complement (G^c)

H_0^{comp} = The genes in G are at most as often DE as the genes in G^c

- Self-contained null

compare gene set to a fixed standard (does not compare to set's complement)

H_0^{self} = No genes in G are DE

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Comparing null hypotheses

- Competitive Null:

- relinquishes power in order to make a stronger statement
- singleton gene sets treated differently from single gene tests
- cannot be used to test set of all genes

- Self-contained Null:

- more statistical power due to restrictive nature (will tend to reject null more often)
- will reject null when gene set is a singleton
- can be used to test set of all genes – useful as precheck

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Possible P-value calculation strategies

■ Gene sampling

genes are sampling units;
permute gene labels – (col. & row sums constant)
(gene set: yes/no; DE: sig/no)

■ Subject sampling

subjects (arrays) are sampling units;
permute array labels (trt/ctrl)

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Comparing resampling strategies

■ Gene sampling

- urn model (like hypergeometric)
 - have two measurements describing each gene: yes/no, sig/no
 - look at all possible re-arrangements of table values, keeping fixed marginal totals
- reverses roles of samples and genes for testing
 - traditional: sample of subjects with fixed measurements
 - here: sample of measurements from fixed samples
- assumes observations for genes are iid
- sample size = number of genes

■ Subject sampling

- sample size = number of samples

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Resampling strategies and P-values

- If we were to replicate the experiment many times (and null is true), P-value is expected % of replicated experiments yielding more extreme results than the actual biological experiment

■ Gene resampling strategy

- a replicate experiment would measure new genes on the same subjects
- does not mimic actual biological experiment
- strongly discouraged

■ Subject resampling strategy

- a replicate experiment would measure new subjects on the same genes

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Hypergeometric test: what does “significance” mean?

- competitive null, gene sampling
- Null assumes genes in gene set are not unusually differentially expressed, and genes in the gene set are independent
- A “significant” P-value → reject Null
 - usually – treat this as concluding “genes in gene set are unusually DE”
 - but – could be due to dependence of genes in gene set (which is to be expected among functionally-related genes)

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Other gene set testing methods

- GSEA (Gene Set Enrichment Analysis) [Excellent 6570 project!]
 - Mootha et al. Nature Genetics 34(3):267-273 (2003)
 - Subramanian et al. PNAS 102(43):15545-15550 (2005)
- Global Ancova
 - Mansmann and Meister. Methods of Information in Medicine 44(3):449-453 (2005)
- SAFE (Significance Analysis of Function and Expression)
 - Barry et al. Bioinformatics 21(9):1943-1949 (2005)
- Global testing
 - Goeman et al. Bioinformatics 20(1):93-99 (2004)
- ADGO (Analysis of Differentially expressed gene sets using composite GO annotation)
 - Nam et al. Bioinformatics 22(18):2249-2253 (2006)
- GXNA (Gene eXpression Network Analysis)
 - Nacu et al. Bioinformatics 23(7):850-858 (2007)
- more ... a very active area right now

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Global test for groups of genes

- Do subjects with similar gene expression profiles have similar class labels?
- Look within single groups of genes
 - GO term
 - KEGG pathway
 - cluster from cluster analysis

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Notation

- n samples (arrays), p genes
- m = subgroup size (# of genes in gene set)
- $X = (x_{ij})$
= matrix of “normalized” expression values
(n rows, p columns)
- Y = vector of “clinical outcome” (usually 0/1)

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Motivation for global test

- Suppose a gene set can be used to predict the clinical outcome
- Then within the gene set, changes in gene expression patterns should match changes in clinical outcome
- Y depends on X – but how?

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Statistical justification for global test

- Generalized linear model:

$$h(E[Y_i | \beta]) = \alpha + \sum_{j=1}^m x_{ij} \beta_j = \alpha + r_i$$

(could be logit function)

- Does gene set predict clinical outcome?

$$H_0 : \beta_1 = \beta_2 = \dots = \beta_m = 0$$

- But – it could be that $m \gg n$
- then classical tests fail

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Revising model as random effects

- Assume: $\beta_1, \beta_2, \dots, \beta_m \sim F$ (generic dist'n)

$$E[\beta] = 0, \text{Var}[\beta] = \tau^2$$

- Then: $H_0 : \tau^2 = 0$

- Equivalently: $r = \begin{bmatrix} r_1 \\ \vdots \\ r_n \end{bmatrix}$, $E[r] = 0$, $\text{Cov}[r] = \tau^2 XX^T$

- Interpretation: $\tau^2 > 0$: similar $X_{i,\bullet}$ and $X_{\ell,\bullet}$
 \Rightarrow similar $E[Y_i]$ and $E[Y_\ell]$

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Score test – from random effects

- Let $R = \frac{1}{m} XX^T$

$$\mu = h^{-1}(\alpha) = E[Y | \tau^2 = 0]$$

$$\mu_2 = E[(Y - \mu)^2 | \tau^2 = 0]; \mu_4 = E[(Y - \mu)^4 | \tau^2 = 0]$$

$$Q = \frac{(Y - \mu)^T R (Y - \mu)}{\mu_2}$$

$$E[Q] = \text{trace}(R); \text{Var}[Q] = 2\text{trace}(R^2) + \left(\frac{\mu_4}{\mu_2^2} - 3\right) \sum_i R_{ii}^2$$

$$c = \text{Var}[Q] / (2E[Q]); v = 2(E[Q])^2 / \text{Var}[Q]$$

- Under $H_0 : \tau^2 = 0$, $Q/c \sim \chi_v^2$

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Global test – interpretation of significance

- Testing a set of genes

Null: none of the genes in the gene set are correlated with clinical outcome

- “For a significant result it is not necessary that the genes in the [gene set] have similar expression patterns, only that many of them are correlated with the [clinical] outcome.”

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```
library(affy); library(ALL); data(ALL); library(genefilter)
eset <- exprs(ALL)
T.cell <- c(rep(0,95),rep(1,33))
```

```
Eset <- new("ExpressionSet", exprs=eset)
pData(Eset) <- data.frame(trt=as.character(T.cell))
annotation(Eset) <- "hgu95av2"
```

```
library(globaltest)
gt.all <- gt(trt,Eset)
# this tests whether B-cell and T-cell
# patients have the same overall gene expression profiles
gt.all
```

	p-value	Statistic	Expected	Std.dev	#Cov
1	2.1e-34	9.98	0.787	0.226	12625

Interpretation: Strong evidence that at least some genes are correlated with T-cell status

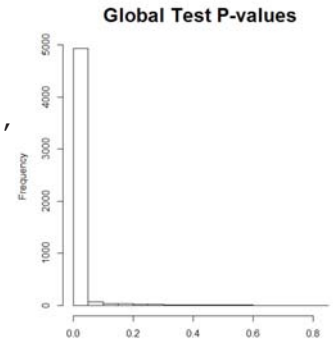
NOTE: In general, use non-filtered data for gene set tests

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```
print(date())
gt.GO <- gtGO(trt, Eset, ontology="BP",
  minsize=10, maxsize=2000)
print(date()) # about 3.5 minutes
```

```
result <- data.frame(GO.ID=names(gt.GO),
  alias=gt.GO@extra[,2],
  pvalue=gt.GO@result[,1])
head(result)
```

```
hist(result$pvalue, xlab=NA,
  main='Global Test P-values',
  cex.main=2)
```



```
dim(result) # 5133 3
```

GO.ID	alias	pvalue
GO:0033077	T cell differentiation in thymus	3.710824e-73
GO:0045061	thymic T cell selection	1.367810e-72
GO:0001775	cell activation	1.403571e-72
GO:0043368	positive T cell selection	1.825101e-71
GO:0045058	T cell selection	2.686312e-71
GO:0046649	lymphocyte activation	1.215176e-70

NOTE: ontology, minsize, and maxsize options in gtGO function – why?

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Global Test – more

- We focus here on logistic regression, but can also do linear regression, multinomial logistic regression, Poisson regression, and Cox proportional hazards model – depending on nature of phenotype variable
- Generalized linear models: Goeman et al. (2011) *Biometrika* 98(2):381-390 – possible 6570 project
- Multiple hypothesis testing – still a problem; but can adjust (even accounting for structure among GO terms – see package vignette)

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Summary

- Testing Gene Sets
 - GO classes
 - KEGG pathways
 - (also Broad Institute pathway databases)
- Some methods based on 2-way table
 - best if based on subject sampling scheme (and self-contained null) – like global testing
- Coming up next: visualization of gene set testing results
- Later: meta-analysis approaches to gene set testing (allow more meaningful alternative)

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